

This article was downloaded by:

On: 30 January 2011

Access details: *Access Details: Free Access*

Publisher *Taylor & Francis*

Informa Ltd Registered in England and Wales Registered Number: 1072954 Registered office: Mortimer House, 37-41 Mortimer Street, London W1T 3JH, UK



Spectroscopy Letters

Publication details, including instructions for authors and subscription information:

<http://www.informaworld.com/smpp/title~content=t713597299>

Dipeptide α -Seryl- β -Methionine—Linear Dichroic IR and Theoretical Characterization

A. Chapkanov^a; S. Zareva^b

^a Department of Chemistry, South-West University, Blagoevgrad, Bulgaria ^b Department of Analytical Chemistry, Faculty of Chemistry, University of Sofia, Sofia, Bulgaria

Online publication date: 17 March 2010

To cite this Article Chapkanov, A. and Zareva, S.(2010) 'Dipeptide α -Seryl- β -Methionine—Linear Dichroic IR and Theoretical Characterization', *Spectroscopy Letters*, 43: 2, 79 — 83

To link to this Article: DOI: 10.1080/00387010903278234

URL: <http://dx.doi.org/10.1080/00387010903278234>

PLEASE SCROLL DOWN FOR ARTICLE

Full terms and conditions of use: <http://www.informaworld.com/terms-and-conditions-of-access.pdf>

This article may be used for research, teaching and private study purposes. Any substantial or systematic reproduction, re-distribution, re-selling, loan or sub-licensing, systematic supply or distribution in any form to anyone is expressly forbidden.

The publisher does not give any warranty express or implied or make any representation that the contents will be complete or accurate or up to date. The accuracy of any instructions, formulae and drug doses should be independently verified with primary sources. The publisher shall not be liable for any loss, actions, claims, proceedings, demand or costs or damages whatsoever or howsoever caused arising directly or indirectly in connection with or arising out of the use of this material.

Dipeptide *L*-Seryl-*L*-Methionine—Linear Dichroic IR and Theoretical Characterization

A. Chapkanov¹
and S. Zareva²

¹Department of Chemistry,
South-West University,
Blagoevgrad, Bulgaria

²Department of Analytical
Chemistry, Faculty of Chemistry,
University of Sofia, Sofia,
Bulgaria

ABSTRACT The dipeptide *L*-seryl-*L*-methionine is elucidated structurally by means of solid-state linear dichroic infrared spectroscopy of oriented samples as a colloidal suspension in nematic liquid crystal as a part of our systematic study of OH-containing small peptides. Vibrational analysis supports the experimental data as well. Quantum chemical *ab initio* calculations are performed in order to obtain the electronic structure and vibrational characteristics of the system studied, where a *cisoid*-configuration of the C=O–NH amide fragment is predicted.

KEYWORDS *ab initio* calculations, linear dichroic infrared spectroscopy, *L*-seryl-*L*-methionine, solid-state

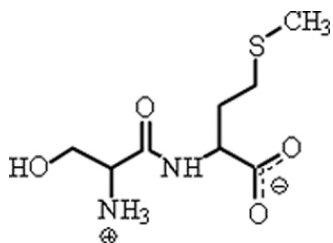
INTRODUCTION

The interaction of Au(III) and Pt(II) with peptides and proteins has been widely investigated in light of the potential antitumor effect of their corresponding coordination compounds.^[1–9] In order to understand the *in vivo* manner and coordination mechanism of these ions with DNA, a systematic examination of their coordination ability with small peptides is necessary.^[1–14] The donor ability of the thioester groups in the methionine side chains is of particular interest. IR spectroscopy and nuclear magnetic resonance methods are powerful tools for studying these interactions, but the comparison of the spectroscopic properties of the pure ligands and their complexes requires knowledge of the corresponding spectroscopic characteristics. However, peptides possess complicated spectroscopic IR patterns requiring the application of specific, unconventional analytical techniques. The possibility of linear-polarized IR spectroscopy (linear dichroic infrared [IR-LD]) or oriented solid samples as a colloidal suspension in nematic liquid crystal address these difficulties to a significant degree. This method has been applied to peptide systems, their complexes, and salts.

A study of the *L*-seryl-*L*-methionine (*Ser-Met*) dipeptide (Scheme 1) is justified, and this article focuses on a preliminary structural analysis of the neutral ligand. In this analysis, theoretical *ab initio* calculations and solid-state IR-LD spectroscopy of oriented colloid suspensions have been applied.

Received 17 May 2008;
accepted 14 May 2009.

Address correspondence to
A. Chapkanov, Department of
Chemistry, South-West University,
“Neofit Rilski,” 66 Ivan Mihailov Str.,
2700 Blagoevgrad, Bulgaria. E-mail:
chapkanov@swu.bg



SCHEME 1 Chemical diagram of *Ser-Met*.

MATERIALS AND METHODS

IR spectra were measured using a Bomem Michelson 100 FT-IR Spectrometer (ABB Bomem, Canada) ($4000\text{--}400\text{ cm}^{-1}$, 2.0 cm^{-1} resolution, 150 scans) equipped with a Perkin Elmer (Waltham, MA, USA) wire-grid polarizer. Nonpolarized solid-state IR spectra were recorded using the potassium bromide disk technique. The oriented samples in the IR-LD spectroscopic study were obtained as colloidal suspensions in a nematic liquid crystal (MLC 6815, Merck, Darmstadt, Germany) with the presence of an isolated nitrile stretching IR band at about 2230 cm^{-1} , which served as an orientation indicator. This solid-state orientation method was used for IR-LD spectroscopy and has been validated.^[15] It uses a colloidal suspension in nematic liquid crystal, which allows accuracy and precision in measuring the influence of the liquid crystal medium on peak positions and integral absorbances of the guest molecule bands. The experimental conditions and design were optimized for the quantitative evaluation of the impact of the four input factors.^[15–17] The number of scans, the rubbing out of KBr plates, the quantity of studied compounds included in the liquid crystal medium, and the ratios of Lorentzian to Gaussian peak functions in the curve-fitting procedure on the spectroscopic signal at five different frequencies were studied. The procedure for determining the position (ν_i) and integral absorbencies (A_i) for each i peak were carried out by deconvolution and curve-fitting procedures at 50:50% ratio of Lorentzian to Gaussian peak functions (χ^2 factors within 0.00066–0.00019 and 2000 iterations). The two treatments are compared using the student t -test. The applicability of this approach of experimentally assigning IR-spectroscopic bands and of obtaining stereostructural information has been demonstrated in a series of organic systems and coordination complexes (e.g., heterocyclic, Cu(II) complexes,

polymorphs, codeine derivatives, peptides their Au(III) complexes, hydrochlorides, and hydrogensquarates^[18–23]). The nature and balance of the forces in the nematic liquid crystal suspension system, the mathematical model for their clearance, the morphology of the suspended particles, and the influence of the space system types on the degree of orientation (i.e., ordering parameter) have been previously shown^[24] using five liquid crystals and 15 compounds. The theory of IR-LD spectroscopy and the use of polarized IR spectra interpretation difference-reduction procedure are given in the reference literature.^[12–24] Quantum chemical calculations were performed with the GAUSSIAN 98 program package.^[25] The output files are visualized using the ChemCraft program.^[26] The dipeptide *Ser-Met* is a Bachem Organics (Budendorf, Switzerland) product.

RESULTS AND DISCUSSION

This work can be considered as a part of systematic spectroscopic and structural elucidation on small peptides, their responsibility to form metal complexes, and their application as potential anticancer medications.^[12–24,27,28]

Theoretical Calculations

The geometries are optimized using two levels of theory: the second-order Moller-Pleset perturbation theory (MP2) and the restricted Hartree-Fock (RHF) theory using a 6-311++G** basis set. The conformational analysis shows several conformers with relative energies (E_{rel}) of less than 5 kJ/mol (Fig. 1) for the zwitterionic and protonated forms of studied peptides. The 3D graph dependence of the E_{rel} as a function of Psi and Phi and angles for the compound studied is shown in Fig. 1, where Psi and Phi describe torsion angles around the $C\alpha\text{--}C$ bond ($\text{Psi}(\psi)\text{--NH}_3^+\text{--CH--C=O}$) and bond $\text{N--C}\alpha$ ($\text{Phi}(\phi)\text{--O=C--NH--CH}$), respectively, in this case.

For the purposes of the experimentally determining structure in solid state, this was used only with the E_{rel} value of 4.1 kJ/mol, the geometry of which is depicted in Fig. 2. The perpendicular orientation of the transition moments were established (Fig. 2).

The RHF approximation gives sufficient comparatively structural parameters for protonated peptides: a conclusion valid for other similar systems as

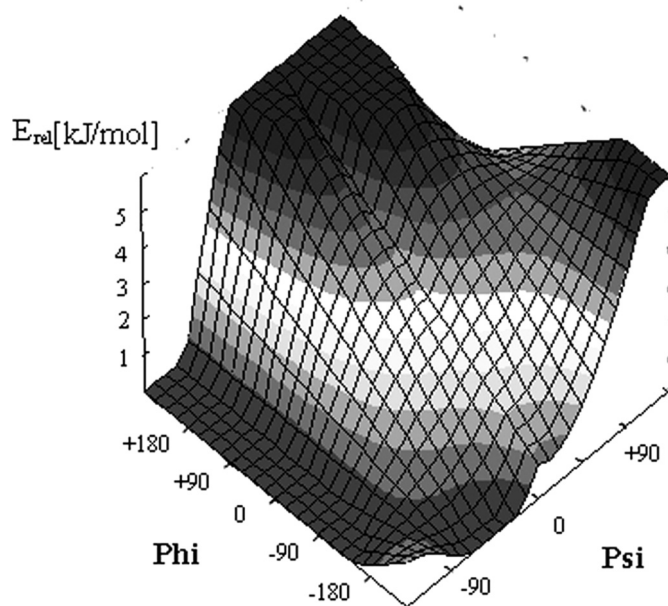


FIGURE 1 3D graph of E_{rel} (kJ/mol) as a function of Phi and Psi angles for *Ser-Met*.

well.^[18–23] The molecular geometry of the studied dipeptide was fully optimized using the force gradient method employing Bernys' algorithm. For every structure, the stationary points found on the molecule potential energy hyper-surfaces were characterized by using standard analytical harmonic vibrational analysis. The absence of imaginary frequencies confirms that the stationary points correspond to the minima of the potential energy hyper-surfaces. The calculation of vibrational

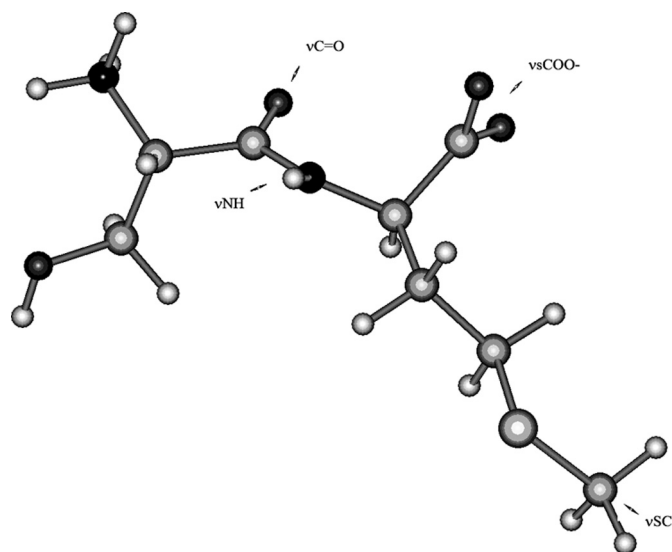


FIGURE 2 Most stable conformer of *Ser-Met* and visualization of selected transition moments.

frequencies and infrared intensities were checked to determine which of the performed calculations agreed well with the experimental data. In this case, the RHF method provided enough accurate vibrational data with calculated standard deviations of 11 cm^{-1} (RHF) and 13 cm^{-1} (MP2), respectively. Therefore, the RHF/6-311++G** data are presented. In order to improve the correspondence between the experimental and theoretical values, the results were modified using an empirical scaling factor of 0.8929.

IR and IR-LD Spectral Analysis

The suspended particles in the nonpolarized IR and difference IR-LD spectra of *Ser-Met* (Fig. 3) showed a significant degree of orientation. The observed pairs of maxima at 3207 cm^{-1} and 3070 cm^{-1} correspond to the stretching ν_{NH} and ν_{OH} modes of the NH and OH groups of the dipeptide molecules that participate in very strong hydrogen bonding. The multiple $3100\text{--}2300\text{ cm}^{-1}$ peak corresponds to the asymmetric and symmetric stretches $\nu_{\text{NH}_3^+}^{\text{as}}$ and $\nu_{\text{NH}_3^+}^{\text{s}}$ vibrations of protonated NH_3^+ groups in the zwitterionic dipeptide. In the $1700\text{--}1400\text{ cm}^{-1}$ range, maxima at 1687 cm^{-1} ($\nu_{\text{C=O}}$, Amide I), 1610 cm^{-1} ($\delta_{\text{NH}_3^+}^{\text{as}}$, NH_3^+ -asymmetric bending vibration), 1563 cm^{-1} ($\nu_{\text{COO-}}^{\text{as}}$, asymmetric stretching mode), 1482 cm^{-1} (δ_{NH} , Amide II), 1502 cm^{-1} ($\delta_{\text{NH}_3^+}^{\text{s}}$), and 1396 cm^{-1} ($\nu_{\text{COO-}}^{\text{s}}$) are observed. The obtained data correlated well with the known theoretical IR spectral for pure amino acids; in methionine, $\delta_{\text{NH}_3^+}^{\text{as}}$ and $\delta_{\text{NH}_3^+}^{\text{s}}$ are observed at 1625 cm^{-1} and 1475 cm^{-1} ,^[21–24,26,29] while in glycine, the $\nu_{\text{COO-}}^{\text{as}}$ and $\nu_{\text{COO-}}^{\text{s}}$ modes are at 1575 cm^{-1} and 1417 cm^{-1} . In contrast to L-threonyl-L-methionine dipeptides^[30] the δ_{NH} is observed in the low-frequency region, which is typical for *cis*- or *cisoid*-configured O=C-NH amide fragments.^[31] A direct experimental confirmation of this assumption is seen in the difference IR-LD spectrum (Fig. 3), where the elimination of the δ_{NH} band and reduction, but not the disappearance of the $\nu_{\text{C=O}}$ (Amide I) mode, is observed. This result is in accordance with the theoretically predicted value of the amide O=C-NH dihedral angle of $163.2(1)^\circ$ (Fig. 2). On the other hand, the spectrum in Fig. 3 is characterized by the not elimination of the $\nu_{\text{COO-}}^{\text{as}}$ and $\nu_{\text{COO-}}^{\text{s}}$ bands, which is impossible in the frame of a structural COO- fragment, due to the

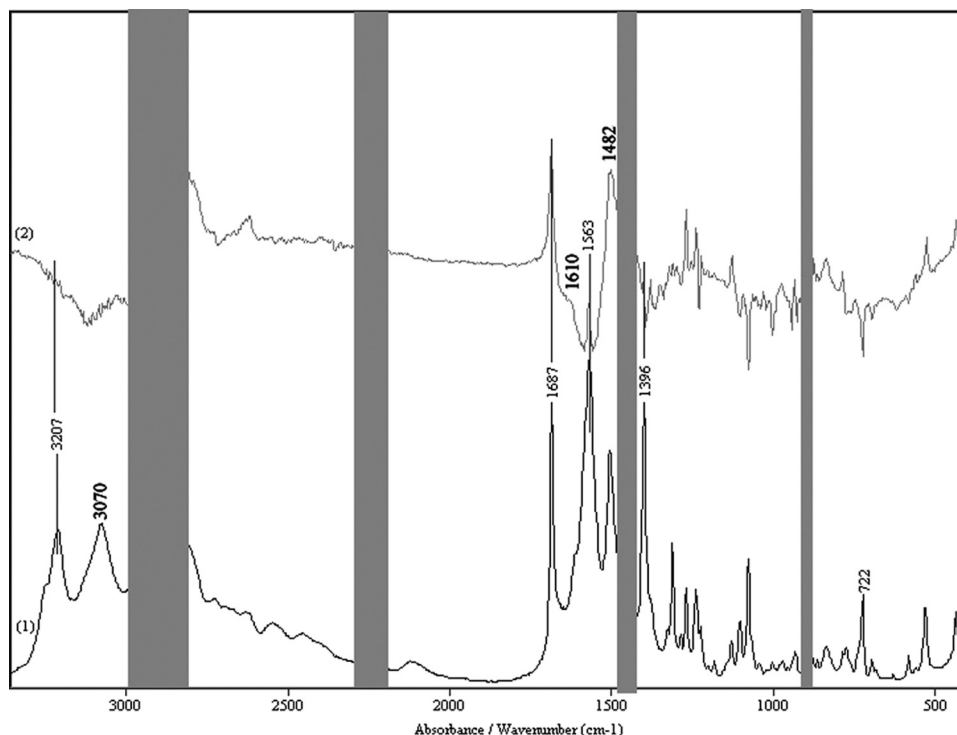


FIGURE 3 Nonpolarized IR (Line (1)) and difference IR-LD (Line (2)) spectra of *Ser-Met*; self-absorption of the liquid crystal is shown with gray rectangles.

perpendicular orientation of the discussed transition moments (Fig. 2). This indicates that the unit cell of *Ser-Met* contains mutually perpendicular oriented molecules as a result of the co-linear orientation of the transition moments in the frame of the neighboring disposed molecules. This phenomenon has been observed in a series of small peptides studied with polarized IR-LD spectroscopy,^[32–34] where the data space group $P2_12_12_1$ (obtained by single crystal X-ray diffraction) is characterized by perpendicularly oriented molecules in the unit cell.

Previous data related to methionine containing tripeptides shows peaks corresponding to νCS .^[14,18–23,29] In pure methionine, the peak is obtained at 725 cm^{-1} .^[29] The elimination of the band at 722 cm^{-1} does not affect the IR characteristic bands and cannot be obtained experimentally, thus providing no proof about its origin. However, it was theoretically predicted that the νCS mode should be observed at 720 cm^{-1} , which means that the band at 722 cm^{-1} corresponds to the stretching νCS vibration.

CONCLUSION

The IR spectroscopic and structural elucidation of the dipeptide *Ser-Met* was performed using IR-LD

spectroscopy of oriented colloid suspensions in nematic host, as well as *ab initio* calculations at RHF/6-311++G** and MP2/6-311++G** levels of theory and basis set. Conclusions about the geometry and IR characteristic bands of the dipeptide were obtained. In contrast to *Thr-Met*, where a *trans*-configuration of the C=O–NH amide fragment is preferable,^[30] in *Ser-Met*, a *cisoid*-configuration is predicted (with a dihedral angle of $163.2(1)^\circ$).

ACKNOWLEDGMENTS

The authors thank Prof. B. Koleva (Sofia University St. Kl. Ohridski) for helpful discussion and for the use of the Dalton 2.0 program package. We also wish to thank Prof. T. Kolev (Bulgarian Academy of Sciences) for the use of the Gaussian 98 program package.

REFERENCES

1. Komeda, S.; Lutz, A.; Spek, L.; Yamanaka, Y.; Sato, T.; Chikuma, M.; Reedijk, J. A novel isomerization on interaction of antitumor-active azole-bridged dinuclear platinum(II) complexes with 9-ethylguanine. Platinum(II) atom migration from N2 to N3 on 1,2,3-triazole. *J. Am. Chem. Soc.* **2002**, *124*(17), 4738–4746.
2. Reedijk, J. Improved understanding in platinum antitumor chemistry. *Chem. Commun.* **1996**, *7*, 801–806.

3. Reedijk, J. Why does *cis*-platin reach guanine-N7 with competing S-donor ligands available in the cell? *Chem. Rev.* **1999**, *99*, 2499–2512.
4. Lippert, B., Ed. *Cisplatin: Chemistry and Biochemistry of a Leading Anticancer Drug*; Wiley-VCH, Weinheim: New York, 1999.
5. Stella v Boom, S.; Chen, B.; Teuben, J.; Reedijk, J. Platinum-thioether bonds can be reverted by guanine-N7 bonds in Pt(dien)²⁺ model adducts. *Inorg. Chem.* **1999**, *38*(7), 1450–1455.
6. Yang, Y.; Tu, C.; Zhang, J.; Lin, L.; Zhang, X.; Liu, O.; Ding, J.; Xu, X.; Guo, N. Novel Au(III) complexes of aminoquinoline derivatives: Crystal structure, DNA binding and cytotoxicity against melanoma and lung tumor cells. *J. Chem. Soc. Dalton Trans.* **2003**, *17*, 3419–3429.
7. Zutphen, S.; Reedijk, J. Targeting platinum anti-tumour drugs: Overview of strategies employed to reduce systemic toxicity. *Coord. Chem. Rev.* **2005**, *249*(24), 2845–2853.
8. Veer, J.; Marel, G.; Elst, H.; Reedijk, J. The antitumor drug *cis*-diamminedichloro platinum(II) preferentially chelating neighboring guanines in the trinucleotide d (pGpGpG. *Inorg. Chem.* **1987**, *26*(14), 2272–2275.
9. Best, S.; Chattopadhyay, T.; Djuran, M.; Palmer, R.; Sadler, P.; Sovago, I.; Varnagy, M. Gold(III) and palladium(II) complexes of glycyl-glycyl-L-histidine: crystal structures of [Au^{III}(Gly-Gly-L-His-H₂)] Cl × H₂O and [Pd^{II}(Gly-Gly-L-His-H₂) × 1.5H₂O and HisNH deprotonation. *J. Chem. Soc. Dalton Trans.* **1997**, 2587–2596.
10. Wienken, M.; Lippert, B.; Zangrando, M.; Randaccio, L. Cold(III) glycyl-L-histidine dipeptide complexes. Preparation and x-ray structures of monomeric and cyclic tetrameric species. *Inorg. Chem.* **1992**, *31*(11), 1983–1985.
11. Wienken, M.; Kiss, A.; Sovago, I.; Fusch, E.; Lippert, B. Ternary palladium(II)–glycylmethionine–nucleobase complexes: Solution studies and crystal structure of the 9-methylguanine compound. *J. Chem. Soc. Dalton Trans.* **1997**, *4*, 563–568.
12. Ivanova, B.; Arnaudov, M.; Todorov, S. Linear-dichroic infrared and NMR spectral analysis of Au³⁺-complex with the tripeptide glycyl-methionyl-glycine. *J. Coord. Chem.* **2006**, *59*, 1749–1755.
13. Ivanova, B. Solid state linear-dichroic infrared spectral analysis of dipeptide *L*-Phe-*L*-Phe and its mononuclear Au(III)-complex. *J. Coord. Chem.* **2006**, *58*, 587–592.
14. Ivanova, B.; Miteva, M. Au(III) interaction with methionine and histidine containing peptides. *J. Coord. Chem.* **2004**, *57*, 217–223.
15. Ivanova, B.; Tsalev, D.; Arnaudov, M. Validation of reducing-difference procedure for the interpretation of non-polarized infrared spectra of n-component solid mixtures. *Talanta* **2006**, *69*, 822–836.
16. Ivanova, B.; Arnaudov, M.; Bontchev, P. Linear-dichroic infrared spectral analysis of Cu(I)-homocysteine complex. *Spectrochim. Acta* **2004**, *60*(4), 855–861.
17. Ivanova, B.; Simeonov, V.; Arnaudov, M.; Tsalev, D. Linear-dichroic infrared spectroscopy—a validation and experimental design of the orientation technique as suspension in nematic liquid crystal. *Spectrochim. Acta* **2007**, *67A*, 66–75.
18. Ivanova, B. Stereo-structural and IR-spectral characterization of histidine containing dipeptides by means of solid state IR-LD spectroscopy and ab initio calculations. *J. Mol. Struct.* **2006**, *782*, 122–126.
19. Ivanova, B.; Arnaudov, M. Solid state linear-dichroic infrared spectral and theoretical analysis of methionine containing tripeptides. *Spectrochim. Acta* **2006**, *65A*, 56–61.
20. Ivanova, B. IR-LD spectroscopic characterization of *L*-tryptophan containing dipeptides. *Spectrochim. Acta* **2006**, *64A*, 931–938.
21. Ivanova, B.; Kolev, T.; Zareva, S. IR-LD spectroscopy characterization of glycyl-containing tripeptides and their hydrochlorides. *Biopolymers* **2006**, *82*, 587–593.
22. Kolev, T.; Ivanova, B.; Zareva, S. Au(III)-complex with dipeptide glycyl-serine—linear polarized IR-spectroscopic, ¹H- and ¹³C-magnetic resonance characterization. *J. Coord. Chem.* **2006**, *60*, 109–115.
23. Kolev, T. Solid-state IR-LD spectroscopic and theoretical analysis of arginine-containing peptides. *Biopolymers* **2006**, *83*, 39–45.
24. Koleva, B.; Kolev, T.; Simeonov, V.; Spassov, T.; Spiteller, M. Linear polarized IR-spectroscopy of partial oriented solids as a colloid suspension in nematic liquid crystal—new tool for structural elucidation of the chemical compounds. *J. Inclusion Phenom.* **2008**, *61*, 319–328.
25. Frisch, M. J.; Trucks, G. W.; Schlegel, H. B.; Scuseria, G. E.; Robb, M. A.; Cheeseman, J. R.; Zakrzewski, V. G.; Montgomery, Jr., J. A.; Stratmann, R. E.; Burant, J. C.; Dapprich, S.; Millam, J. M.; Daniels, A. D.; Kudin, K. N.; Strain, M. C.; Farkas, Ö.; Tomasi, J.; Barone, V.; Cossi, M.; Cammi, R.; Mennucci, B.; Pomelli, C.; Adamo, C.; Clifford, S.; Ochterski, J.; Petersson, G. A.; Ayala, P. Y.; Cui, Q.; Morokuma, K.; Salvador, P.; Dannenberg, J. J.; Malick, D. K.; Rabuck, A. D.; Raghavachari, K.; Foresman, J. B.; Cioslowski, J.; Ortiz, J. V.; Baboul, A. G.; Stefanov, B. B.; Liu, G.; Liashenko, A.; Piskorz, P.; Komáromi, I.; Gomperts, R.; Martin, R. L.; Fox, D. J.; Keith, T.; Al-Laham, M. A.; Peng, C. Y.; Nanayakkara, A.; Challacombe, M.; Gill, P. M. W.; Johnson, B.; Chen, W.; Wong, M. W.; Andres, J. L.; Gonzalez, C.; Head-Gordon, M.; Replogle, E. S.; Pople, J. A. *Gaussian 98*; Gaussian, Inc.: Pittsburgh, PA, 1998.
26. Zhurko, G. A.; Zhurko, D. A. ChemCraft: Tool for treatment of chemical data, Lite version build 08 (freeware), 2005.
27. Zareva, S.; Kolev, T.; Chapkanov, A.; Spiteller, M.; Koleva, B. Solid-state spectroscopic and structural elucidation of the dipeptide *L*-phenylalanyl-*L*-alanine dihydrate (*H*-Phe-Ala-OH·2H₂O) and its hydrochloride (*H*-Phe-Ala-OH·HCl). *Polish J. Chem.* **2009**, *83*, 421–430.
28. Chapkanov, A.; Miteva, Y.; Kolev, T.; Spiteller, M.; Koleva, B. New Au(III), Pt(II) and Pd(II) complexes with pentapeptide glycyl-glycyl-L-methionyl-glycyl-glycine and their interaction with calf thymus DNA. *Prot. Peptide Lett.* **2010**, *17*(2), 228–237.
29. Cao, X.; Fischer, G. Conformational and infrared spectral studies of L-methionine and its N-deuterated isotopomer as isolated zwitterions. *J. Phys. Chem. A* **2002**, *106*(1), 41–50.
30. Chapkanov, A.; Zareva, S. Structural and IR-spectroscopic elucidation of dipeptide *L*-threonyl-*L*-methionine in solid-state. *Prot. Peptide Lett.* **2009**, *16*(11), 1277–1280.
31. Arnaudov, M.; Ivanova, B.; Todorov, S.; Zareva, S. Reducing-difference infrared spectral analysis of *cis*- and *trans*-configured lactams. *Spectrochim. Acta, Part A* **2006**, *63*, 491–500.
32. Ivanova, B.; Kolev, T.; Zareva, S. Solid-state IR-LD spectroscopic and theoretical analysis of glycine-containing peptides and their hydrochlorides. *Biopolymers* **2006**, *82*, 587–596.
33. Koleva, B.; Kolev, T.; Spiteller, M. Structural and spectroscopic analysis of hydro-gensquarates of glycine-containing tripeptides. *Biopolymers* **2006**, *83*, 498–507.
34. Kolev, T. Solid-state IR-LD spectroscopic and theoretical analysis of arginine-containing peptides. *Biopolymers* **2006**, *83*, 39–45.